

This product is for research use only (not for diagnostic or therapeutic use)

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Product no AS12 2110

Anti-PIP2-1-7 | Plasma membrane aquaporin isoforms 1-7, C-terminal

Product information

Immunogen

KLH-conjugated synthetic peptide derived from Zea mays PIP2-7 C-terminal, Q9ATM4, conserved also in Zea mays PIP2-1, UniProt: Q84RL7, PIP2-2, UniProt: Q9ATM8, PIP2-3 (80 % conservation) UniProt: Q9ATM7, PIP2-4 (80 % conservation) UniProt: Q9ATM6, PIP2-5 (70 % conservation) UniProt: Q9XF58, PIP2-6 (50 % conservation) UniProt: Q9ATM5

Host Rabbit

Clonality Polyclonal

Purity Serum

Format Lyophilized

Quantity 50 μl

Reconstitution For reconstitution add 50 µl of sterile water

Storage

Store lyophilized/reconstituted at -20°C; make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of

Application information

Recommended dilution 1 : 600 (IP), 1: 3000 (WB)

Expected | apparent

30.7 | 30 kDa (Zea mays)

Confirmed reactivity Lactuca sativa, Pisum sativum, Solanum lycopersicum, Zea mays

Predicted reactivity

Arabidopsis thaliana, Artemisia annua, Brassica oleracea, Capsicum annuum, Capsicum chinense, Cicer arietinum, Coffea arabica, Cucumis melo, Cucumis sativus, Fragaria chiloensis, Glycine max, Helianthus annuus, Hordeum vulgare, Malus prunifolia, Medicago trunculata, Mimosa pudica, Nicotiana tabacum, Noccaea caerulescens, Olea europaea, Oryza sativa, Phaseolus vulgaris, Pisum sativum, Prunus mume, Pyrus communis, Spinacia oleracea, Solanum lycopersicum, Solanum tuberosum, Trifolium repens, Triticum urartu, Triticum aestivum, Vitis vinifera Species of your interest not listed? Contact us

Not reactive in Allium sativum, Hordeum vulgare

Additional information

Detection pattern consists of di and monomer of PIP2-7.

This antibody has a potential to work in immunolocalization studies, as it is recognizing C-terminal part of the sequence.

This product can be sold containing ProClin if requested.

Selected references

Paluch-Lubawa and Polcyn (2024). Tissue-specific accumulation of PIP aquaporins of a particular heteromeric composition is part of the maize response to mycorrhiza and drought. Sci Rep. 2024 Sep 17;14(1):21712.doi: 10.1038/s41598-024-72828-8.

Kumar et al. (2024). Dehydration-responsive cytoskeleton proteome of rice reveals reprograming of key molecular pathways to mediate metabolic adaptation and cell survival. Plant Physiol Biochem. 2024 Feb:207:108359. Kumar et al. (2022). Proteomic dissection of rice cytoskeleton reveals the dominance of microtubule and microfilament proteins, and novel components in the cytoskeleton-bound polysome, Plant Physiology and Biochemistry, Volume 170,2022, Pages 75-86, ISSN 0981-9428, https://doi.org/10.1016/j.plaphy.2021.11.037.

Application example



10 µg of total protein from Zea mays roots (1), Phaseolus vulgaris leaves (2) or roots (3) extracted with a mixture of 250 mM sorbitol, 50 mM Tris-HCl (pH 8), 2 mM EDTA, and protease inhibitors [1 mM phenylmethylsulfonyl Xuoride, 1 mg ml⁻¹ each of leupeptin, aprotinin, antipain, chymostatin, and pepstatin were separated on 12 % SDS-PAGE and blotted 1h to PVDF. Blots were blocked with 5% milk in TBS-T for 2h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 3.000 for 1h at RT with agitation. The antibody



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solution was decanted and the blot was rinsed briefly twice, then washed four times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (goat anti-rabbit IgG horse radish peroxidase conjugated, from Agrisera AS09 602) diluted to 1:30 000 in TBS-T for 1h at RT with agitation. The blot was washed as above and developed for 5 min with ECL according to the manufacturers instructions. Exposure time was 60 seconds.

Courtesy of Dr. Ricardo Aroca, CSIC, Spain